

(12) INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

(19) World Intellectual Property
Organization
International Bureau



549991

(43) International Publication Date
30 September 2004 (30.09.2004)

PCT

(10) International Publication Number
WO 2004/083820 A2

(51) International Patent Classification⁷: **G01N**
(21) International Application Number:
PCT/US2004/008558

GB, GR, HU, IE, IT, LU, MC, NL, PL, PT, RO, SE, SI, SK,
TR), OAPI (BF, BJ, CF, CG, CI, CM, GA, GN, GQ, GW,
ML, MR, NE, SN, TD, TG).

(22) International Filing Date: 19 March 2004 (19.03.2004)

Declarations under Rule 4.17:

(25) Filing Language: English

(26) Publication Language: English

(30) Priority Data:
60/455,970 19 March 2003 (19.03.2003) US

(71) Applicant (for all designated States except US):
TRUSTEES OF BOSTON UNIVERSITY [US/US];
One Sherborn Street, Boston, MA 02215 (US).

(72) Inventors; and

(75) Inventors/Applicants (for US only): **UNLU, Selim, M.**
[US/US]; 2 Jamaicaaway Court, Jamaica Plain, MA 02130
(US). **BERGSTEIN, David, A.** [US/US]; 87 Brighton
Avenue, Allston, MA 02134 (US). **RUANE, Michael, F.**
[US/US]; 16 Corey Road, Brookline, MA 02245 (US).
GOLDBERG, Bennett, B. [US/US]; 26 Berkshire Road,
Newton, MA 02460 (US).

(74) Agents: **GAGNEBIN III, Charles, L.** et al.; Weingarten,
Schurgin, Gagnebin & Lebovici, LLP, Ten Post Office
Square, Boston, MA 02109 (US).

(81) Designated States (unless otherwise indicated, for every
kind of national protection available): AE, AG, AL, AM,
AT, AU, AZ, BA, BB, BG, BR, BW, BY, BZ, CA, CH, CN,
CO, CR, CU, CZ, DE, DK, DM, DZ, EC, EE, EG, ES, FI,
GB, GD, GE, GH, GM, HR, HU, ID, IL, IN, IS, JP, KE,
KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MA, MD,
MG, MK, MN, MW, MX, MZ, NA, NI, NO, NZ, OM, PG,
PH, PL, PT, RO, RU, SC, SD, SE, SG, SK, SL, SY, TJ, TM,
TN, TR, TT, TZ, UA, UG, US, UZ, VC, VN, YU, ZA, ZM,
ZW.

(84) Designated States (unless otherwise indicated, for every
kind of regional protection available): ARIPO (BW, GH,
GM, KE, LS, MW, MZ, SD, SL, SZ, TZ, UG, ZM, ZW),
Eurasian (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), Euro-
pean (AT, BE, BG, CH, CY, CZ, DE, DK, EE, ES, FI, FR,

— as to applicant's entitlement to apply for and be granted
a patent (Rule 4.17(ii)) for the following designations AE,
AG, AL, AM, AT, AU, AZ, BA, BB, BG, BR, BW, BY, BZ,
CA, CH, CN, CO, CR, CU, CZ, DE, DK, DM, DZ, EC, EE,
EG, ES, FI, GB, GD, GE, GH, GM, HR, HU, ID, IL, IN, IS,
JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MA,
MD, MG, MK, MN, MW, MX, MZ, NA, NI, NO, NZ, OM,
PG, PH, PL, PT, RO, RU, SC, SD, SE, SG, SK, SL, SY, TJ,
TM, TN, TR, TT, TZ, UA, UG, UZ, VC, VN, YU, ZA, ZM,
ZW, ARIPO patent (BW, GH, GM, KE, LS, MW, MZ, SD,
SL, SZ, TZ, UG, ZM, ZW), Eurasian patent (AM, AZ, BY,
KG, KZ, MD, RU, TJ, TM), European patent (AT, BE, BG,
CH, CY, CZ, DE, DK, EE, ES, FI, FR, GB, GR, HU, IE, IT,
LU, MC, NL, PL, PT, RO, SE, SI, SK, TR), OAPI patent (BF,
BJ, CF, CG, CI, CM, GA, GN, GQ, GW, ML, MR, NE, SN,
TD, TG)

— as to the applicant's entitlement to claim the priority of the
earlier application (Rule 4.17(iii)) for the following desig-
nations AE, AG, AL, AM, AT, AU, AZ, BA, BB, BG, BR, BW,
BY, BZ, CA, CH, CN, CO, CR, CU, CZ, DE, DK, DM, DZ,
EC, EE, EG, ES, FI, GB, GD, GE, GH, GM, HR, HU, ID,
IL, IN, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU,
LV, MA, MD, MG, MK, MN, MW, MX, MZ, NA, NI, NO, NZ,
OM, PG, PH, PL, PT, RO, RU, SC, SD, SE, SG, SK, SL, SY,
TJ, TM, TN, TR, TT, TZ, UA, UG, UZ, VC, VN, YU, ZA,
ZM, ZW, ARIPO patent (BW, GH, GM, KE, LS, MW, MZ,
SD, SL, SZ, TZ, UG, ZM, ZW), Eurasian patent (AM, AZ,
BY, KG, KZ, MD, RU, TJ, TM), European patent (AT, BE,
BG, CH, CY, CZ, DE, DK, EE, ES, FI, FR, GB, GR, HU, IE,
IT, LU, MC, NL, PL, PT, RO, SE, SI, SK, TR), OAPI patent
(BF, BJ, CF, CG, CI, CM, GA, GN, GQ, GW, ML, MR, NE,
SN, TD, TG)

— of inventorship (Rule 4.17(iv)) for US only

Published:

— without international search report and to be republished
upon receipt of that report

For two-letter codes and other abbreviations, refer to the "Guid-
ance Notes on Codes and Abbreviations" appearing at the begin-
ning of each regular issue of the PCT Gazette.

(54) Title: **RESONANT CAVITY BIOSENSOR**

(57) Abstract: An assay system having a channel bounded by first and second reflective surfaces adapted to accommodate a fluid material therebetween and defining a plurality of regions in an array between those surfaces with each region defining a resonant cavity and adapted to receive a capturing material on a surface thereof whereby a source of radiation illuminates each region to provide a standing wave of radiation of within the cavity indicative of binding of said capturing agent to material under investigation, a binding thereof being detected in response to radiation from each cavity indicative of a change in the standing wave pattern.

WO 2004/083820 A2

TITLE OF THE INVENTION
RESONANT CAVITY BIOSENSOR

CROSS REFERENCE TO RELATED APPLICATIONS

5 This application claims priority to commonly assigned
Provisional Application Serial No. 60/445,970 filed March 19,
2003. International Application No. PCT/US00/12287, filed
10 05 May 2000 is incorporated herein by reference.

STATEMENT REGARDING FEDERALLY SPONSORED RESEARCH OR
DEVELOPMENT

N/A

BACKGROUND OF THE INVENTION

15 In bio-research, ecological testing, medical testing, drug
testing and bio-weapon and hazards detections there is a need
20 for rapid, simultaneous and real time detection of various
agents. Typically the agents are provided in a fluid to a test
structure, generally an array of some form in which capturing
materials of diverse types capable of binding to one or more of
the materials undergoing test and provided in the fluid medium.
25 The binding is a result of an affinity that molecules or bio-
molecules have for each other and includes the affinities of
DNA, RNA, proteins, small molecules and other molecules. DNA
arrays and protein arrays, commonly called DNA or protein chips,
are two technologies used for bio-molecule affinity sensing in
30 such fields as genomics and proteomics.

 The array of capturing materials is created in a known
pattern such that by correlation of the binding response of the
capturing material to the fluid born molecules under test, it is

possible by detecting the level of binding at each array element to determine what materials under test are present. In one case of DNA or RNA testing, various sequences of the DNA or RNA molecule are affixed to corresponding locations in the array.
5 DNA or RNA in the fluid being tested will tend to bind where the sequences therein strongly match the sequences attached to the various array sites.

Test methods known to date fail to provide high throughput, real time operations or to avoid difficult labeling processes, or to avoid capturing material incompatibility with metal sensor
10 surface, requiring cumbersome linking chemistry that may adversely affect binding properties.

Among the techniques previously used which fail to provide all of these requirements in combination are fluorescent
15 tagging. Fluorescent tagging procedures suffer from a number of problems including the difficulty of tagging and the possibility of tagging altering the binding properties. Moreover tagging procedures are difficult to monitor continuously in real time. Among other techniques surface plasmon resonance is popular.
20 This technique, however, requires the affixation of molecules to a metal surface, particularly gold, which has the above mentioned incompatibility problem. Other techniques include waveguide techniques and acoustic detection techniques. Neither of these accommodates a high throughput, requiring a large
25 number of array elements.

Finally, another known technique, reflectometric interference spectroscopy, suffers from the complication of using multiple fiber probes, which greatly hinders its ability to become high throughput.

30

BRIEF SUMMARY OF THE INVENTION

The present invention provides an affinity detection system which has high throughput, is real time and avoids the

complexities of metal, cumbersome equipment and other deficiencies of prior art techniques.

According to the teaching of the present invention, an array of biosensor or capturing material elements is formed between first and second surfaces of reflective mirrors. Typically, the mirrors are formed by multi-layer dielectric surfaces selected to be reflective at a particular range of wavelengths, commonly in the IR frequency regions. Light, typically from a laser, is applied through the array and focused onto a photo-detection system such as a CCD chip or photodetector array where each element of the capturing array is focused onto one or more pixels of the image chip.

A fluid containing materials under test flows through the array between the surfaces forming the mirror elements, the material having an affinity for a capturing material in one or more array cells. Instead of a fluid (gas or liquid with particles or fluid components under test) a solid of appropriate transparency may be tested. These materials will be bound to the capturing material of that cell having such an affinity changing the resonant properties of the resonant cavity formed between the mirror surfaces in that cell. The result will be a change in the light received by the corresponding CCD pixels. That change can be detected by processing electronics correlating the position of the change, its nature and the known affinity of that particular resonant cavity cell.

In this fashion, a large number of bio-molecules or other molecules can be tested for in real time. Array dimensions of hundreds of thousands or millions of cells are possible and the processing electronics available today can easily provide a real time indication of the nature of molecules present in a medium being tested.

BRIEF DESCRIPTION OF THE SEVERAL VIEWS OF THE DRAWINGS

These and other features of the present invention are described in the following detailed description taken in conjunction with the drawing of which:

Fig. 1 is a diagram illustrating the components of a testing system of the present invention;

Fig. 2 illustrates an array of bio-probes according to the invention;

Fig. 3 illustrates an array of photo-detectors such as a photodetector array for use in the present invention or alternatively as a multi-cell light source according to an embodiment of the invention;

Fig. 4 illustrates the operation of a single cell in response to incident light according to the invention;

Fig. 5 illustrates a standing wave pattern typical of the present invention when exposed to illumination;

Figs. 6A and 6B illustrate the variation in cell sensitivity as a function of dielectric layers in opposing mirrors according to the invention;

Figs. 7A and 7B illustrate the change in resonant wavelength with the cavity narrowing from binding of molecules at different cavity dimensions and mirror compositions;

Figs. 8A, 8B and 8C illustrate the variation of cavity mirror transmittance as a function of wavelength with cavity mirror dielectric layer complexity;

Figs. 9A and 9B illustrate resonance under differing conditions useful in understanding the invention;

Figs. 10A through 10E illustrate an example of the invention in actual test use;

Fig. 11 illustrates an embodiment with an integral photodetector; and

Fig. 12 illustrates a system for testing multiple arrays on a continuous basis.

DETAILED DESCRIPTION OF THE INVENTION

5

Fig. 1 illustrates an optical control system according to the invention in which a wavelength tunable laser 1, which may be an Aristu MNG638A provides output radiation typically in the vicinity of 1560 nanometers (15,600 Angstroms) to a single mode optical fiber 2. Radiation in the fiber 2 is applied to an optical attenuator 3, which may be an Agilent 85156A. The attenuator provides dynamic adjustment consistency to promote the operation of the system as a whole as described below. In order to provide beam cleaning which insures a Gaussian distribution to the spatial intensity profile of the beam, the output of the attenuator 3 is applied through a long coil of single mode fiber 4, typically 5Km. The thus cleaned light is applied through a fiber collimator 5 which uses an antireflective coated objective at 1550 nanometers. The collimator produces a 1mm beam, the diameter being measured between half maximum intensity points.

The collimated beam 6 is applied to an optical system comprising a lens 7 which is in the exemplary example a 15mm focal length antireflective coated lens. The converging beam from the lens 7 is applied through a 50 micron aperture 8 placed at the focal point 9 of the beam from the lens 7. The function of the 50 micron aperture is to provide further beam cleaning. The thus clean beam is applied to a further antireflective coated lens 10, typically of a 125mm focal length. The lens 10 produces a collimated beam 11, in this example of 10mm width between half maximum intensity points. The beam 11 is reflected at right angles by a 45° mirror 12 into the array of cells. This optical system may be a microscope system.

The detection system comprises a cavity formed between first and second reflecting surfaces 14 and 15 separated by a space 16 within which a standing wave is generated by the radiation in beam 11. That radiation is applied through a lower support 17, aperture 18 into a lower or first stage 19 through a further aperture 20.

The second reflective surface 15 is supported by a second stage 21 supported by a second support 22. Micrometers 23 and 24 allow adjustment of the first stage 19 with respect to the first support 17 while a micrometer 25 provides a similar function for the second stage 21 with respect to the support 22.

The light in beam 11 creates a standing wave pattern in a cavity 16, particularly one dependent upon the characteristics of a capturing material applied to each cell in an array on the mirror 14, all as described more completely herein below. Light of an intensity dependant upon the degree of resonance within each cell travels in a beam 30 through a focusing lens 31 to a camera 32, typically by reflection from a 45° mirror 33. The camera 32, which may be a Sensors Unlimited SU128-1.7R camera having a InGaAs sensor with pixels in a 128 x 128 array, receives that light. Typically, each cell will be imaged onto one or more pixels in the array of camera 32. The image from camera 32 is read into a computer 34 to an image acquisition card 35, which may typically be a National Instrument NI-PCI1422. The computer 34 has a controller card, typically a GPIB card, which applies control signals to the tunable laser 1. The card 36 also operates through a piezoelectric controller 37 to control piezoelectric actuators on the adjusters 23 and 24, typically placed at their tips where they joint the stage 19. The computer 34 may have an input/output interface 38 for communication with users, networks, printers display and other typical computer accessories.

The computer maintains a feedback loop through the piezoelectric controller 7 on the actuators 23 and 24 via the camera 32 to sense fringe patterns in the optical image received and processed by the camera 32 which are an indication of an out
5 of parallel condition between the stages 19 and 21, using known minimization techniques, the piezoelectric drives are operated to minimize those fringing elements thereby obtaining a parallel condition of the stages 19 and 21. The piezo elements are also operable by the computer to vary the spacing between reflectors
10 as an alternative or complementary to wavelength scanning of the laser radiation.

A heating element 42 operates with a heat control unit 43 which may or may not have a connection to computer 34 in order to maintain or control the temperature between the stages 19 and
15 21 and in particular within the region 16 where standing wave patterns are created by the incident illumination. This heat control accomplishes the function of avoiding dynamic changes on the mirrors during testing.

The computer 34 is programmed to process the image data
20 from each pixel received by the camera 32 in order to determine the thickness between the reflectors 14 and 15 in each cell, representative of the binding of material flowed through the intermirror region 16 for biologic or chemical assay purposes. This process includes the steps of:

- 25 1. Low pass filtering the intensity wavelength response curves for each pixel with respect to wavelength;
2. Cross-correlating the local overlapping groups of pixels to find relative shifts in the intensity wavelength profile;
- 30 3. Solving an over-determined problem which involves integrating the shifts to find a consistent picture of capturing material surface thickness.

In order to develop an intensity wavelength response, the computer 34 will typically cause the tunable laser 1 to scan through a set of wavelengths.

5 In the operation of the feedback control of mirror alignment, if there is an angle between the two mirrors, such that the distance between them changes by more than a half wavelength, the cavity will be resonant in some places and non-resonant at others. Everywhere the resonance condition is satisfied, the camera and computer will see bright spots on the
10 camera monitoring transmission. For a perfectly flat mirror at an angle, this amounts to horizontal lines indicating equal cavity spacing where resonance is satisfied. As one of the angles is tuned, the lines grow closer together or farther apart. Closer together, indicates that the angle perpendicular
15 to the lines is growing steeper. As one of the adjustment knobs is tuned far to one end, the lines will grow increasing close together and increasing perpendicular to that angle. If the same knob is turned the other direction, the lines grow closer and less perpendicular to that angle. As the knob is kept
20 turning, the lines will go through an optimal position after which they will again start to grow closer together and more perpendicular to the direction of angle change. By adjusting very carefully, one can tune to that optimal position where the lines would start contracting again if there was movement in
25 either direction of the tuning knob, and where the lines are actually parallel to the direction of the angle adjustment. This means that in this direction, the surface is completely flat. The same is repeated for the other direction. Once the other direction is done, the first one has probably moved a bit
30 due to vibration from handling of the system, so an iterative approach may be needed to some extent. Wavelength changes move the lines but not their orientation, and the spacing between lines only changes slightly due to wavelength and can be taken

into account by simply realizing that it was due to wavelength not angle change. Most often, we can not get to the point that the surface is completely lit due to surface curvature. A circle is eventually seen instead of the whole screen going
5 bright, because the surface is curved and satisfying the resonant condition at only places on the circle, no matter how parallel the mirrors are. When this circle is visible, the mirrors are reasonably parallel and tuning can stop.

The piezo / computer control can take over this function and it allows much finer adjustment, making it easier to tune. The piezos don't disturb the system like the hand of a human operator does on the micrometer does. It is then possible to control the system to keep the mirrors parallel throughout the operation of the biosensor.

15 Fig. 2 illustrates an array of cells 80, typically those which may be applied to the surface of mirror 14 in bio or protein chips of known design. Hundreds of thousands or even millions of cells 80 can be provided in the mirror 14 within the channel 10. These cells, as noted above, are typically imaged
20 into one or more pixels 82 of a photodetector array 84 in Fig. 3 within the multi-channel detector 30. Alternatively, the pattern illustrated by Fig. 3 can represent the pattern of light emitters such as from laser diodes. The memory associated with processor 34 will correlate one or more of the pixels 82 of the
25 photodetector array 84 to corresponding cavities 80 and the particular molecular affinity of the material bonded to the mirror 14, typically to several angstroms in depth.

Fig. 4 illustrates diagrammatically the operation of a single cell 80 of the detection system of the present invention.
30 The cell has a thin layer of a capturing material 90 affixed to it at each cell 80. The optical thickness of this material may be 5 angstroms and when binding occurs with molecules for which

the capturing material there has an affinity the optical thickness may increase by as much as 10 angstroms.

Light from the laser 1 provides wavefronts 100 which pass into the cavity 80 through mirror 12 and are reflective within the cavity by the reflectance of the mirrors 14 and 15 to the wavelength and the incident radiation 100. As molecules in the flow through channel 16 bind to the region 90, the wavelength response will shift from an original array 102 in Fig. 4A to a shifted wavelength response 104 in Fig. 4B.

Fig. 5 illustrates in greater detail the mirrors 14 and 15 as consisting of a plurality of alternating silicon and silicon dioxide dielectric layers 106 and 108, respectively. As illustrated in Fig. 5, layer 14 will typically be terminated with an extra silicon dioxide layer 108 causing a standing wave pattern 110 within the cavity illustrated in Fig. 5 to have a peak 112 at the outer wall of the layer 108. This maximizes the effectiveness and sensitivity of the detection system of the present invention.

This effect is more clearly illustrated in Figs. 6A and 6B and accompanying, corresponding Figs. 7A and 7B. Fig. 6A illustrates substantially the standing wave pattern and dielectric layer scheme of Fig. 5. Fig. 7A illustrates the wavelength shift corresponding to a 5 nanometer buildup of material at the corresponding cell.

Fig. 6B illustrates the case where the standing wave pattern is a minimum at the front of the mirror 14 and the corresponding relatively insignificant change in resonant wavelength for a 5 nanometer buildup being illustrated in Fig. 7B.

In Fig. 7A:

Cavity is 50 micron cavity filled with a buffer solution;
Left curve is for 5 Angstroms of biomaterial;
Right curve is for 10 Angstroms of biomaterial;

270nm extra SiO₂ layer is present to maximize sensitivity;
AR coating is present on mirror backsides;
X-axis is wavelength in nm;
Y-axis is transmission;
5 Curves shifted 0.004nm.

In Fig.7B:

Cavity is 50 micron cavity filled with a buffer solution;
Left curve is for 5 Angstroms of biomaterial;
Right curve is for 10 Angstroms of biomaterial;
10 No extra SiO₂ layer is present to maximize sensitivity;
AR coating is present on mirror backsides;
X-axis is wavelength in nm;
Y-axis is transmission;
Curves nearly indistinguishable; Curves shifted 0nm.

15 Figs. 8A and 8C illustrate the sensitivity increase as the
number of dielectric layers within the mirrors 15 and 14
increases essentially forming a Bragg reflector. Figs. 8A-8B
curves represent computer simulations corresponding, from left
to right, to 2 sets of alternating high and low index layers, 3
20 sets, and 4 sets in the far right. The calculations use a
solution index of refraction of 1.33 and the material is
calculated with an index of refraction of 1.45. The capturing
material layer is 0.05 nanometers while the target/capture layer
thickness is 0.1 nanometers.

25 The embodiments of the invention may vary. In particular,
the wavelength of the applied radiation may be other than within
the infrared or IR bands, the radiation applied to the mirrors
15 and 14 may be other than orthogonal and the mirrors 15 and 14
may not necessarily be parallel. The processor and
30 photodetector array, while typically measuring light amplitude
as an indication of affinity binding, may detect phase,
polarization or actual frequency shifts. The tuning of the
laser 36 may be continuous or in discrete steps. A VCSEL array

may alternatively be used as a source of radiation as well as laser diodes.

SIMULATION

5 Figs. 9A-9B show a simulation of wavelength shifts.

In Fig. 9A:

Cavity is 10 micron cavity filled with a buffer solution;

Left curve is for 5 Angstroms of biomaterial;

Right curve is for 10 Angstroms of biomaterial;

10 270nm extra SiO₂ layer is present to maximize sensitivity;

AR coating is present on mirror backsides;

X-axis is wavelength in nm;

Y-axis is transmission;

Curves shifted 0.012nm.

15 In Fig. 9B:

Cavity is 100 micron cavity filled with a buffer solution;

Left curve is for 5 Angstroms of biomaterial;

Right curve is for 10 Angstroms of biomaterial;

270nm extra SiO₂ layer is present to maximize sensitivity;

20 AR coating is present on mirror backsides;

X-axis is wavelength in nm;

Y-axis is transmission;

Curves shifted 0.001nm.

EXAMPLE

25 Fabrication of test sample: A test pattern has been fabricated in SiO₂ to test that the system is working (Fig. 10A). 270nm of SiO₂ was deposited on the top surface of the first reflector. This SiO₂ layer serves to place the sensing surface at a position in the cavity where the field strength is high (approximately 1 quarter wavelength out from the reflector

surface at the wavelengths we are scanning at). The SiO₂ surface was then masked and lightly etched to leave 4 square features. The 4 squares are 50 μ m x 50 μ m and 30 μ m apart. The pattern is repeated every 500 μ m. The sample was masked at the
5 boxes and wet etched everywhere else using HF to remove approximately 15nm of material. 4 boxes 50 μ m x 50 μ m x 15nm should remain on top of 255nm SiO₂, on top of the reflector.

Running experiment: Micrometers were used to position the reflectors close to each other to form the cavity. A z-stage
10 was used to translate one of the reflectors to approximately 100 μ m away from the other, to form a 100 μ m air cavity. Fringes could be seen on the video output from the camera, indicating that the reflectors were not parallel. An angle stage holding the other reflector was then carefully adjusted until the
15 fringes could no longer be seen. Wavelength was scanned from 1545nm to 1560nm in 0.01nm steps. An image of the cavity was captured at each step with approximately 6X magnification (Figs. 10B and 10C out and in resonance).

Processing data: The resulting wavelength response curve
20 for each pixel was then low pass filtered with a 5 samples/nm cutoff. The data was then broken down into groups of 9 waveforms taken from 3x3 sets of neighboring pixels. The groupings were made so that they overlap by 1 row or column of pixels with neighboring groups of 3x3. Within each 3x3 group,
25 the 9 wavelength response curves were cross correlated to each other. The peak of the cross-correlations indicates the shift between those two waveforms. 9 waveforms cross correlating with each other produces 81 correlations, including 9 auto correlations, which leads to 72 shifts describing the relative
30 position of each pixel with respect to the other 8 pixels. This information is heavily redundant. A linear systems over determined problem was setup and solved to find 8 shifts for 8 of the pixels relative to the top left most pixel which was

given the shift of zero. This was done for all of the overlapping groups of 3x3 pixels. The top left most group of 3x3 pixels was designated to have a zero overall offset. The offset of the other 3x3 groupings relative to this first 3x3 group was then determined. The offset for each of the 3x3 groupings was found from solving a linear systems over determined problem as well, where the equations are derived from the fact that the 3x3 groupings overlap by columns and rows that must be consistently the same height. The solution of this problem provided an overall offset for each of the 3x3 groups. The final result is a mesh where the height of each pixel indicates the shift between its wavelength response and that of the upper left most pixel on the camera. There are two key advantages to this technique. First, only local waveforms are ever correlated directly. This is important because the wavelength response drifts in overall shape across the sensor surface due to inhomogeneous illumination and curvature of the mirror structure. Comparing only local pixels, we are more assured that the resonant waveform has the same shape and its only the shift we are measuring. Secondly, by comparing each pixel to 8 of its neighbors, redundancy is gained which is used to improve the accuracy of the observed shift over a correlation done between just two pixels.

Interpreting the results: The four boxes of Fig. 10E in the mesh reflect the results. The boxes appear to be approximately 10 steps high. The steps indicated on the z-axis correspond to the 0.01nm steps in wavelength that were taken.

The best sensitivity we could attain by including the extra SiO₂ layer would be a $2/m$ shift in wavelength for a corresponding shift in surface height where m is the mode number given by $m=2*d/\lambda$, λ being the wavelength and d being the cavity size. For a 100 μ m cavity and wavelengths in the neighborhood of 1.55 μ m, the sensitivity would be 0.0155nm shift

in wavelength response for a 1nm shift in sensor surface height. Alternatively stated, every 1nm shift in wavelength indicates a 65nm ($1/0.0155$) shift in sensor surface height. Again, this assumes a quarter-wavelength of SiO₂ on the reflector surface.
5 As this SiO₂ layer would differ towards a half wave thickness, or zero thickness, the sensitivity would fall to 0.

With this in mind, we see that the 10 steps for the features in the mesh surface plot indicates a 0.1nm shift in wavelength, which indicates a 6.5nm step in the sensor surface.
10 The target height of the features was 15nm.

In Fig 10A:

Model of SiO₂ pattern created by photolithography and wet HF etch.

15 Surface sits on top of quarter wavelength layer of SiO₂ (270nm) for maximized sensitivity.

In Figs. 10B and C:

View from camera at one fixed wavelength ($\lambda=1559.70\text{nm}$). Approximately 6x magnification so that each
20 pixel represents approximately 10 microns square.

At most wavelengths the image is completely dark, and at a few, its nearly all bright. Here, at $\lambda=1559.70\text{nm}$, most of the surface is on its way to resonance (bright), but the 4 apparent squares in the upper right are lagging because of their
25 shifted response.

In Fig. 10D:

Bottom axis shows wavelength in nanometers. Vertical axis indicates relative intensity as measured by camera pixel. Previous image was taken at 1559.70nm where we can see that the
30 intensity was on the rise at this pixel, but not maximum. On the transition, the contrast is maximized and one is able to discern the 4 square features.

Here is the final mesh of shift vs. pixel position. The horizontal axes indicate pixel (this is a 40x40 section taken from the 128x128 array for clarity). The vertical axis indicates by how many steps the response was shifted relative to the bottom most corner pixel which was designated as having a 0 shift. The data was taken in steps of $\lambda = 0.01\text{nm}$, so that the features, which appear to be 10 steps high, corresponds to a 0.1nm shift in wavelength for their response. For this 100 μm cavity, every nanometer shift in resonant response indicates a 65nm shift, so that these features would appear to be 6.5nm in height.

Finally, the processing while still substantially real time may involve other or alternative mathematical techniques such as averaging, differentiating, integrating, curve fitting (in lieu of correlating), or correlating or otherwise comparing various frames or pixels of the multi-channel detector.

Fig. 11 illustrates an embodiment where the photodetector is provided on the exit mirror as an array 140 on a substrate 142, which may be silicon. Here as throughout the optical path an anti-reflective coating 144 is provided between them. Dielectric layers 146 are at the bottom of the substrate to provide the reflectivity described above. The bottom reflector structure 148 as above comprises a substrate 150, bottom coating 152 anti-reflective to the incident collimated beam 154. The substrate 150 has dielectric layers 156 and a silicon dioxide layer 158 on which the array of capturing material 160 is placed.

Fig. 12 is an embodiment where plural testing units such as the bottom reflector structure 148 are passed under the upper reflector structure 162 of the type described above on a conveyor system 160. The light from the upper structure 162 after passing through the cavity 164 test material and capturing

material 166 is received at a detection system 168 which may be as described above.

The invention described herein is to be limited only in accordance with the following claims:

CLAIMS

What is claimed is:

1. An assay system comprising:

5 a channel bounded by first and second reflective surfaces adapted to accommodate therebetween a fluid having material to be tested;

10 a plurality of regions in a pattern of an array between said first and second surfaces, each region defining a resonant cavity and adapted to receive a capturing material on one of the first and second surfaces therein;

a source of radiation to illuminate each region at a wavelength adapted to provide a standing wave of radiation within each said cavity;

15 a detector for the radiation in each said cavity and operative to indicate a change in the standing wave pattern reflective of binding of capturing material with material in a fluid within each said cavity.

20 2. An assay system comprising:

a channel bounded by first and second reflective surfaces adapted to accommodate therebetween a fluid having material to be tested;

25 a plurality of regions in a pattern of an array between said first and second surfaces, each region defining a cavity and adapted to receive a capturing material on one of the first and second surfaces therein;

30 a source of wavelength scanned radiation to illuminate each region at a wavelength adapted to provide a transmission of that radiation within each said cavity representative of material from said fluid bound to said capturing material;

a detector for the radiation in each said cavity and operative to indicate the level of binding by said capturing material of material in said fluid within each said cavity.

- 5 3. The assay system of claim 1 or 2 wherein said first and second reflective surfaces include one or more dielectric layers forming said reflective surface at a wavelength corresponding to said standing wave pattern.
- 10 4. The assay system of any previous claim wherein the capturing material of each respective cavity is of a nature to bind with different materials in the fluid.
- 15 5. The assay system of any previous claim wherein said capturing material as applied to each cavity forms a DNA or protein chip where individual capturing materials in each cavity are DNA or protein selective.
- 20 6. The assay system of any previous claim wherein said radiation source is an IR source.
7. The assay system of any previous claim wherein said radiation source is a laser source.
- 25 8. The assay system of any previous claim wherein said radiation source is a tunable laser source.
- 30 9. The assay system of claim 7 further including means for sweeping the wavelength of said tunable laser over a range including a wavelength corresponding to said standing wave pattern in each cavity.

10. The assay system of any previous claim further including a beam expander in a path of radiation between said radiation source and said channel.
- 5 11. The assay system of any previous claim further including a beam condenser in a path of radiation between said channel and said detector.
- 10 12. The assay system of any previous claim wherein said detector includes a multi element detector wherein each element receives radiation from a corresponding cavity.
13. The assay system of any previous claim wherein said detector is a CCD detector.
- 15 14. The assay system of any previous claim wherein said first and second surfaces are parallel and radiation from said source is applied othogonally to said first and second surfaces.
- 20 15. The assay system of any one of previous claims 1 to 12 wherein said radiation is applied obliquely to at least one of said first and second surfaces.
- 25 16. The assay system of any previous claim wherein said detector detects one or more of radiation amplitude, phase, polarization and wavelength.
- 30 17. The assay system of any one of previous claims 1 to 6 wherein said source of radiation includes means for causing said radiation to emit at discrete different wavelengths.
18. The assay system of any previous claim including means for controlling a temperature of fluid within said channel.

19. The assay system of any previous claim further including means for dynamically varying spacing of said first and second surfaces.

20. The assay system of any previous claim wherein said detection system includes a photodetector array integral with a support for one of said reflective surfaces which is not supporting a capturing material.

21. The assay system of any previous claim wherein said reflective surface having said capturing material thereon has an added dielectric layer to provide a peak in a standing wave pattern in said cavity at said capturing material.

22. The assay system of any previous claim further including means for varying the spacing of said reflective surfaces to vary the cavity resonance condition.

23. A method for assaying comprising:

providing a channel bounded by first and second reflective surfaces adapted to accommodate a fluid material therebetween;

providing a plurality of regions in a pattern of an array between said first and second surfaces, each region defining a resonant cavity and adapted to receive a capturing material on one of the first and second surfaces therein;

applying radiation to illuminate each region at a wavelength adapted to provide a standing wave of radiation within each said cavity;

detecting the radiation in each said cavity and operative to indicate a change in the standing wave pattern reflective of binding of capturing material with material in a fluid within each said cavity.

24. A Method for assaying comprising:

providing a channel bounded by first and second reflective surfaces adapted to accommodate therebetween a fluid having material to be tested;

providing a plurality of regions in a pattern of an array between said first and second surfaces, each region defining a cavity and adapted to receive a capturing material on one of the first and second surfaces therein;

scanning source of radiation applied to illuminate each region at a wavelength adapted to provide a transmission of that radiation within each said cavity representative of material from said fluid bound to said capturing material;

detecting the radiation in each said cavity and operative to indicate the level of binding by said capturing material of material in said fluid within each said cavity.

25. The assay method of claim 23 or 24 wherein said first and second reflective surfaces include one or more dielectric layers forming said reflective surface at a wavelength corresponding to said standing wave pattern.

26. The assay method of any one of previous claims 23 to 25 wherein the capturing material of each respective cavity is of a nature to bind with different materials in the fluid.

27. The assay method of any of previous claims 23 to 26 wherein said capturing material as applied to each cavity is provided as a DNA or protein chip where individual capturing materials in each cavity are DNA or protein selective.

28. The assay method of any of previous claims 23 to 27 wherein said radiation is IR.

29. The assay method of any of previous claims 23 to 27 wherein said radiation is laser radiation.

5 30. The assay method of any of previous claims 23 to 29 including the step of tuning said radiation.

10 31. The assay method of claim 30 further including the step of sweeping the wavelength of said radiation over a range including a wavelength corresponding to said standing wave pattern in each cavity.

15 32. The assay method of any of previous claims 23 to 31 further including the step of expand said radiation in a beam along a path of radiation between said radiation source and said channel.

20 33. The assay method of any of previous claims 23 to 32 further including the step of condensing a beam of radiation along a path of radiation between said channel and said detector.

25 34. The assay method of any of previous claims 23 to 33 wherein said detecting step includes detecting in each of a plurality of detection elements wherein each element receives radiation from a corresponding cavity.

30 35. The assay method of any of previous claims 23 to 34 wherein said first and second surfaces are parallel and radiation from said source is applied othogonally to said first and second surfaces.

36. The assay method of any one of previous claims 23 to 35 wherein said radiation is applied obliquely to at least one of said first and second surfaces.

5 37. The assay method of any of previous claims 239 to 36 wherein said detection step detects one or more of radiation amplitude, phase, polarization and wavelength.

10 38. The assay method of any one of previous claims 23 to 37 wherein said radiation is emitted at discrete, different wavelengths.

15 39. The assay method of any of previous claims 23 to 38 including the step of controlling a temperature of fluid within said channel.

20 40. The assay method of any of previous claims 23 to 39 further including the step of dynamically varying spacing of said first and second surfaces.

25 41. The assaying method of any previous claims 23 through 40 wherein said detecting step includes detecting at a photodetector array integral with a support for one of said reflective surfaces which is not supporting a capturing material.

30 42. The assaying method of any of claims 23 through 41 wherein said reflective surface is provided having said capturing material thereon has an added dielectric layer to provide a peak in a standing wave pattern in said cavity at said capturing material.

43. The assay system of any of claims 23 through 42 further including varying the spacing of said reflective surfaces to vary the cavity resonance conditions.

5 44. An assay system comprising:

a zone bounded by first and second reflective surfaces adapted to accommodate therebetween a material to be tested;

10 a plurality of regions in a pattern of an array between said first and second surfaces, each region defining a resonant cavity between the first and second surfaces therein;

a source of radiation to illuminate each region at a wavelength adapted to provide a standing wave of radiation within each said cavity;

15 a detector for the radiation in each said cavity and operative to indicate a change in the standing wave pattern reflective material within each said cavity.

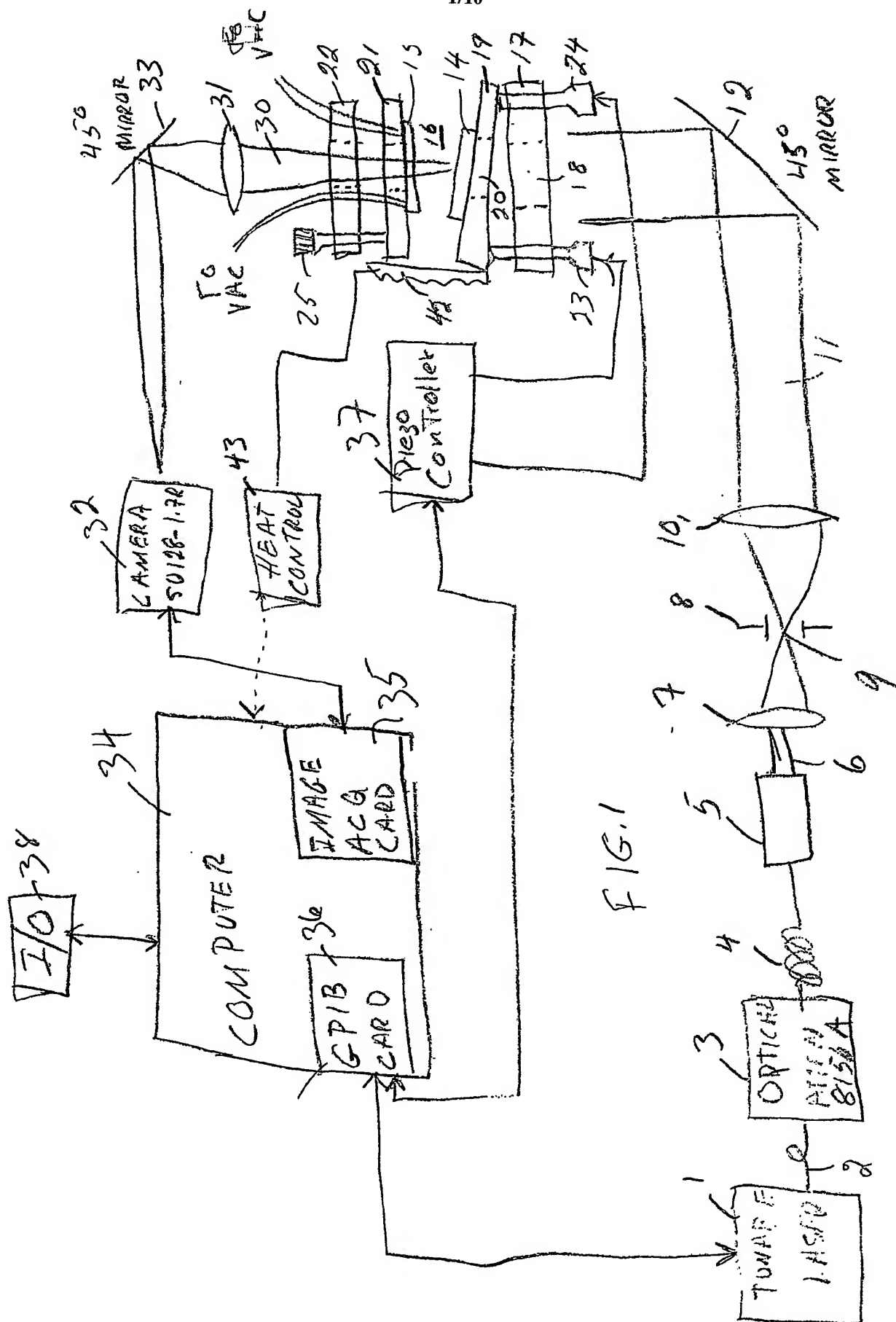
45. An assay system comprising:

20 a channel bounded by first and second reflective surfaces adapted to accommodate therebetween a material to be tested;

a plurality of regions in a pattern of an array between said first and second surfaces, each region defining a cavity between the first and second surfaces therein;

25 a source of wavelength scanned radiation to illuminate each region at a wavelength adapted to provide a transmission of that radiation within each said cavity representative of said material;

30 a detector for the radiation in each said cavity and operative to indicate the level material within each said cavity.



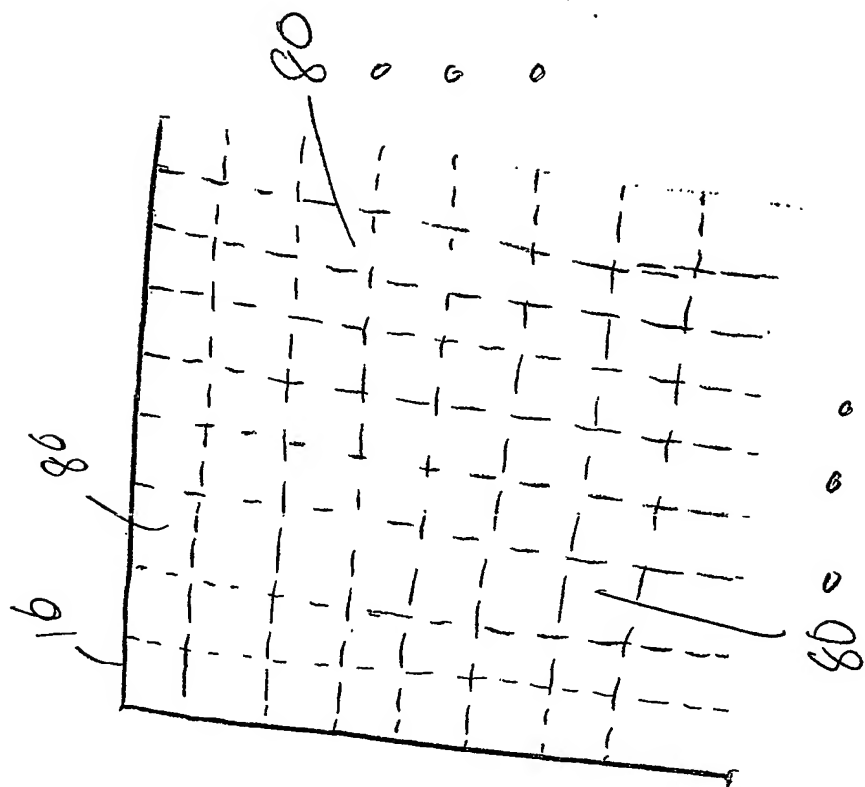


FIG 2

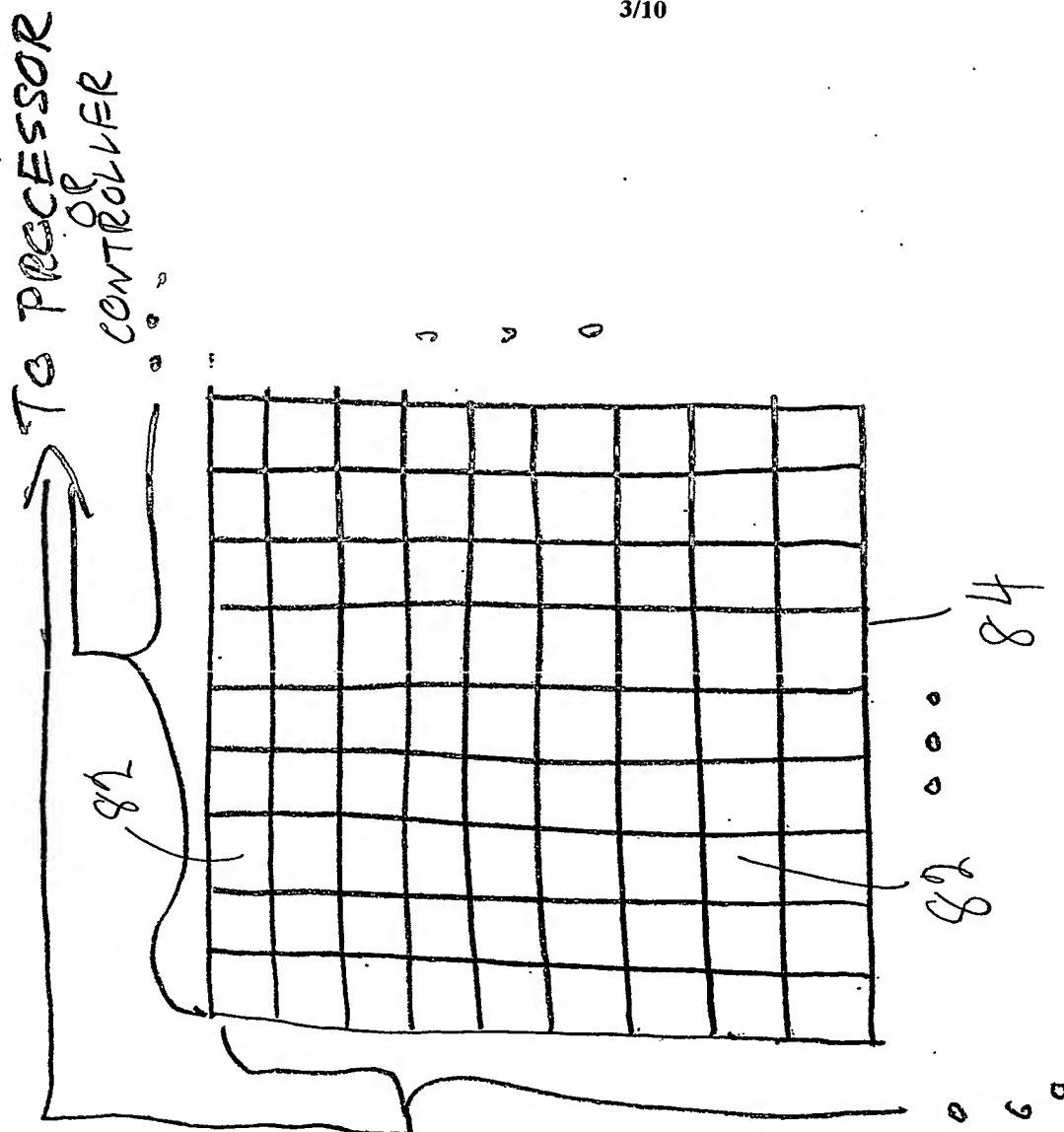
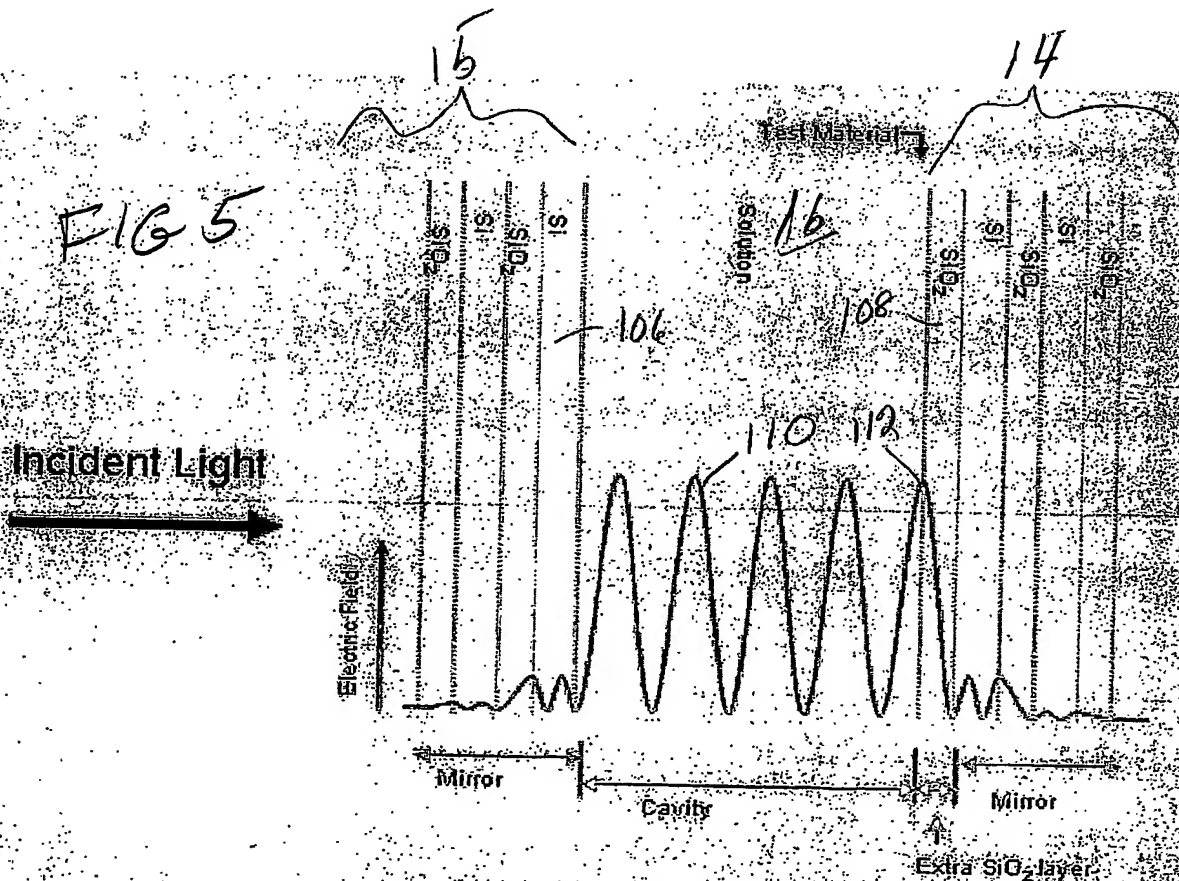
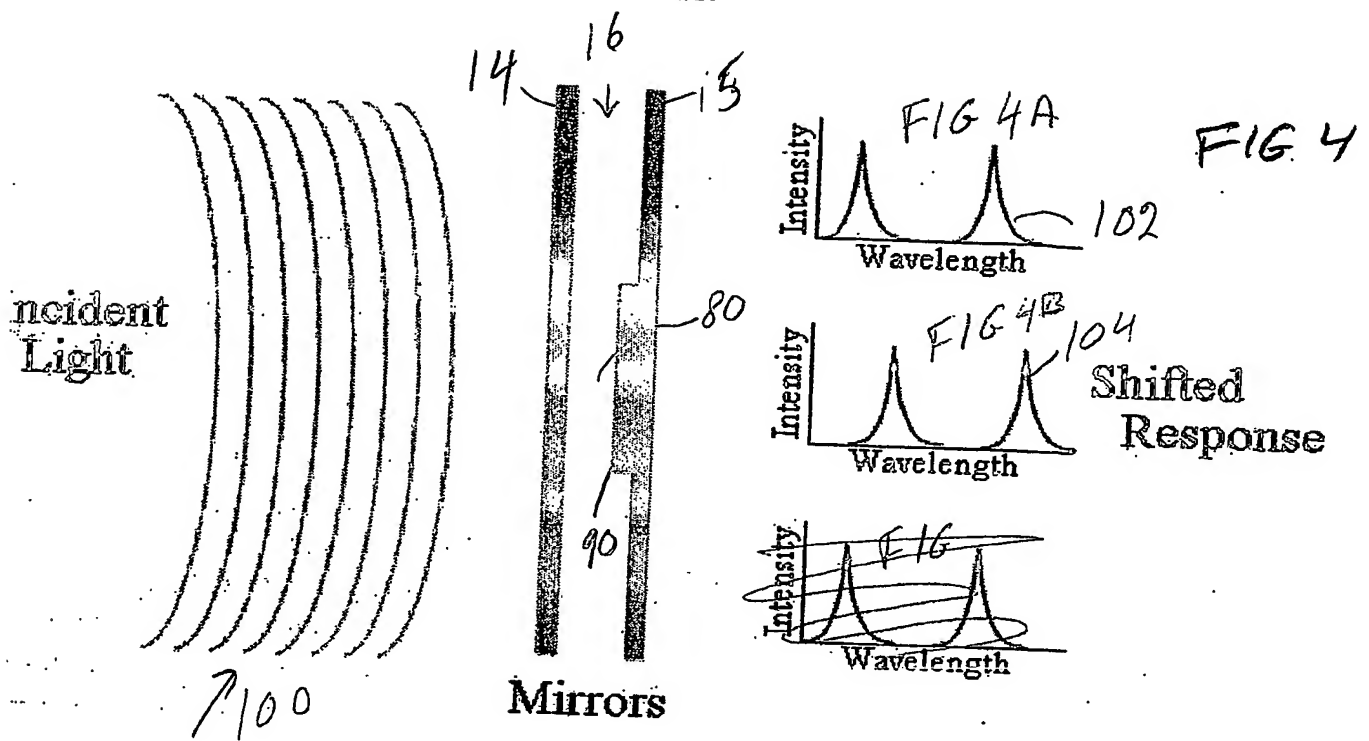


FIG. 3



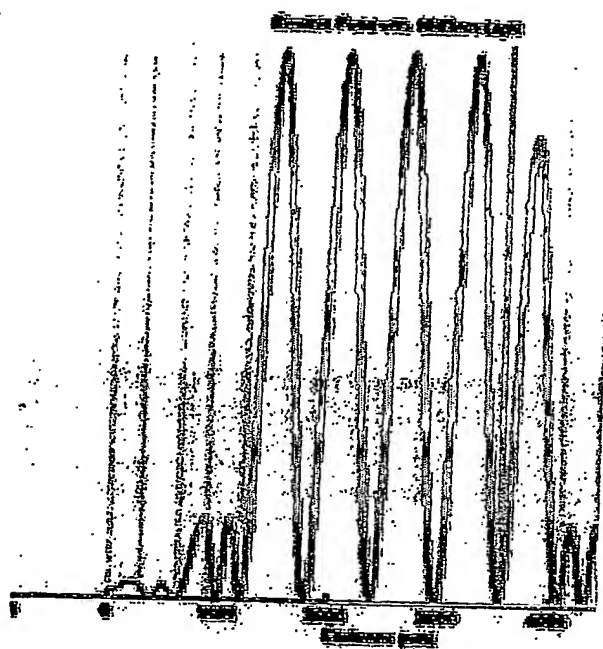


FIG 6B

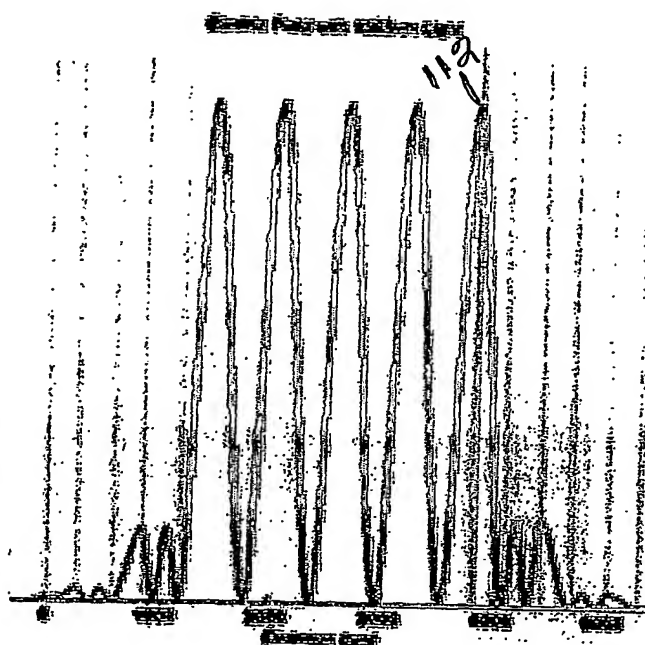


FIG 6A

Fig 7B

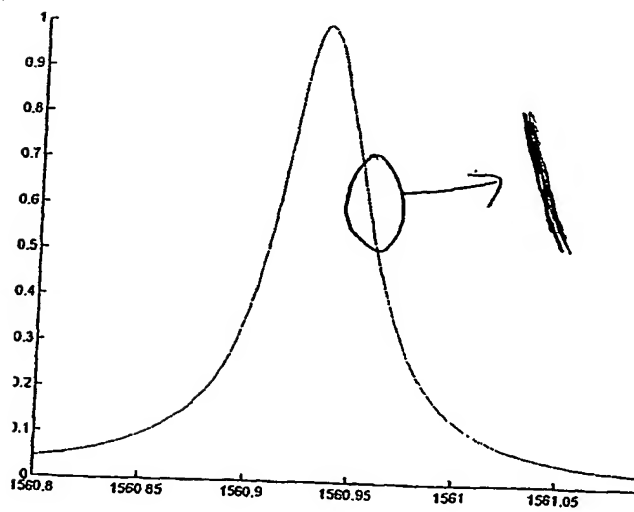
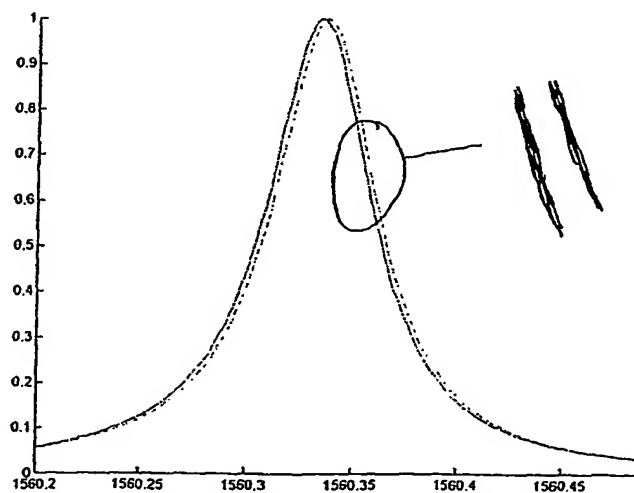
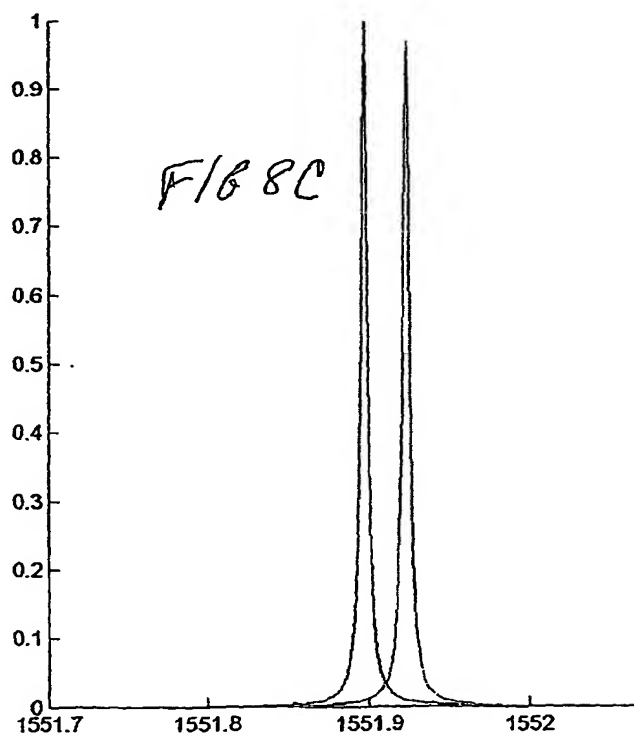
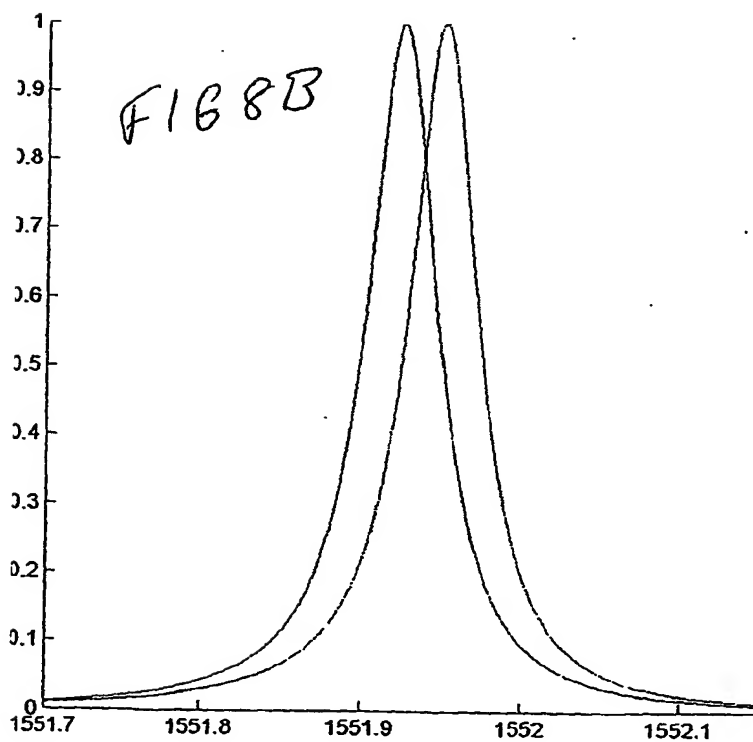
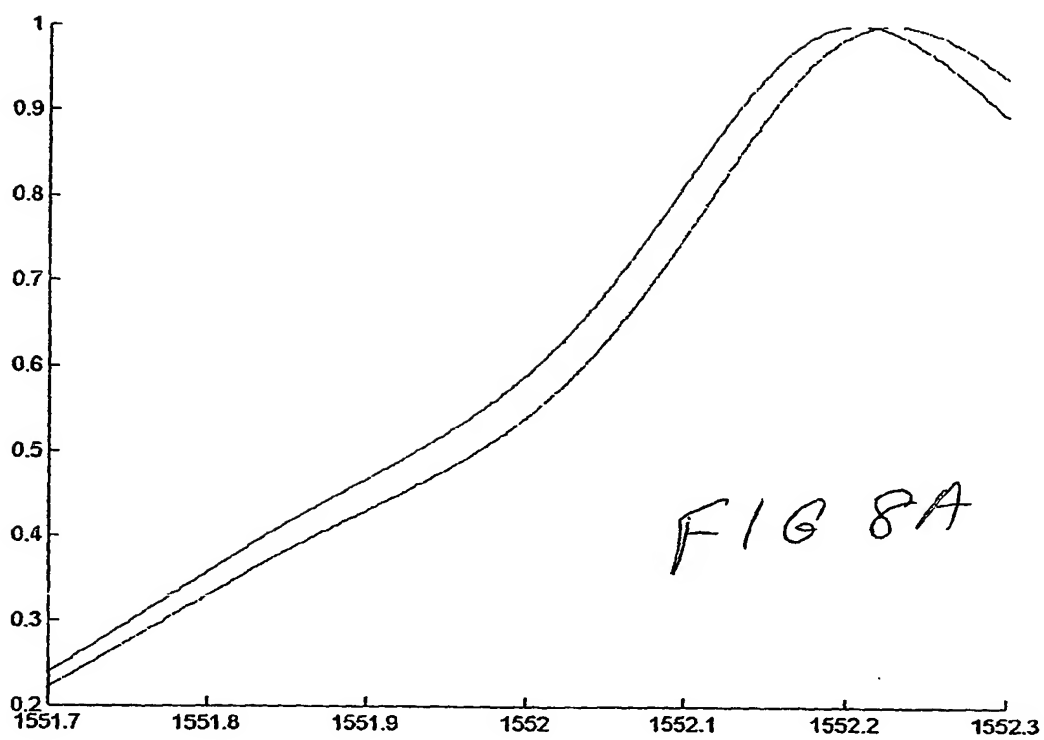


Fig: 7A





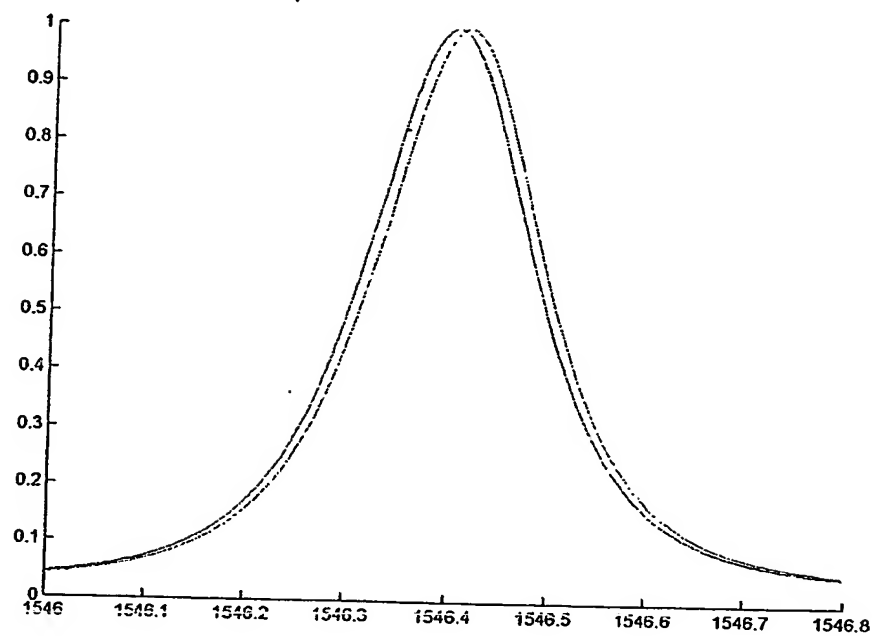
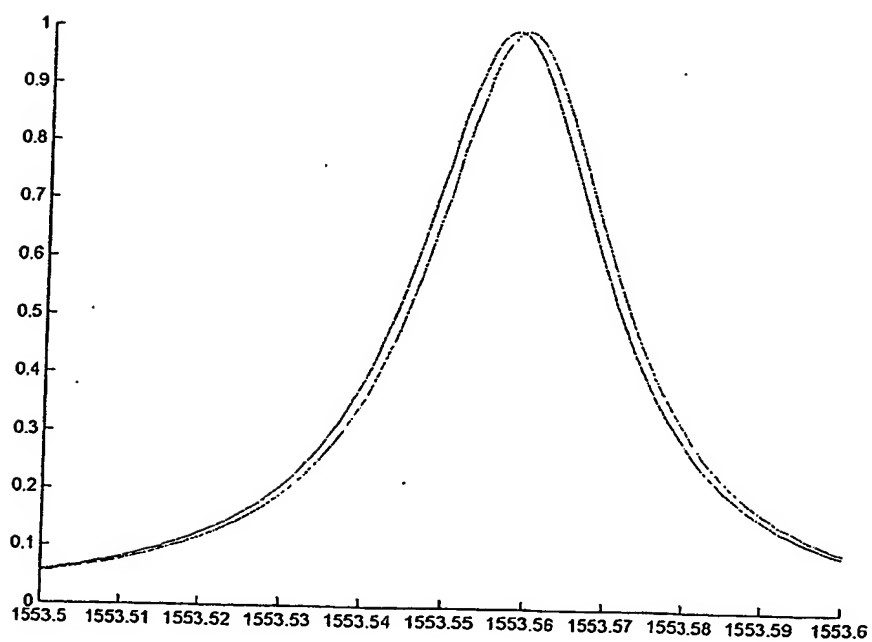


Fig 9
~~10~~ B



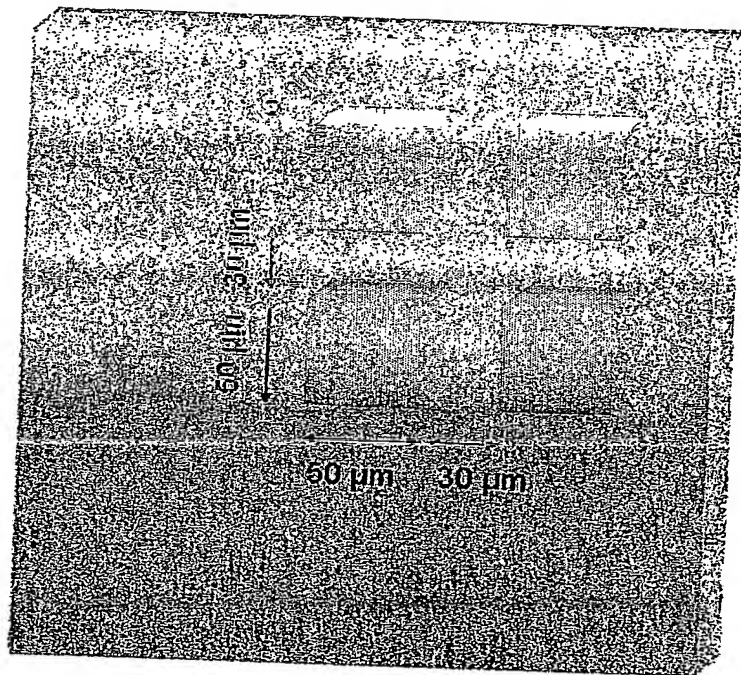


FIG 10A

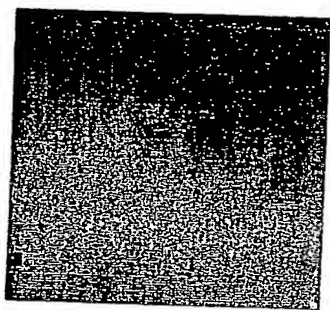


FIG 10B

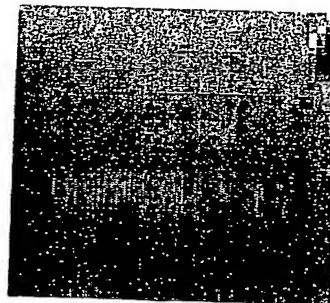


FIG 10C

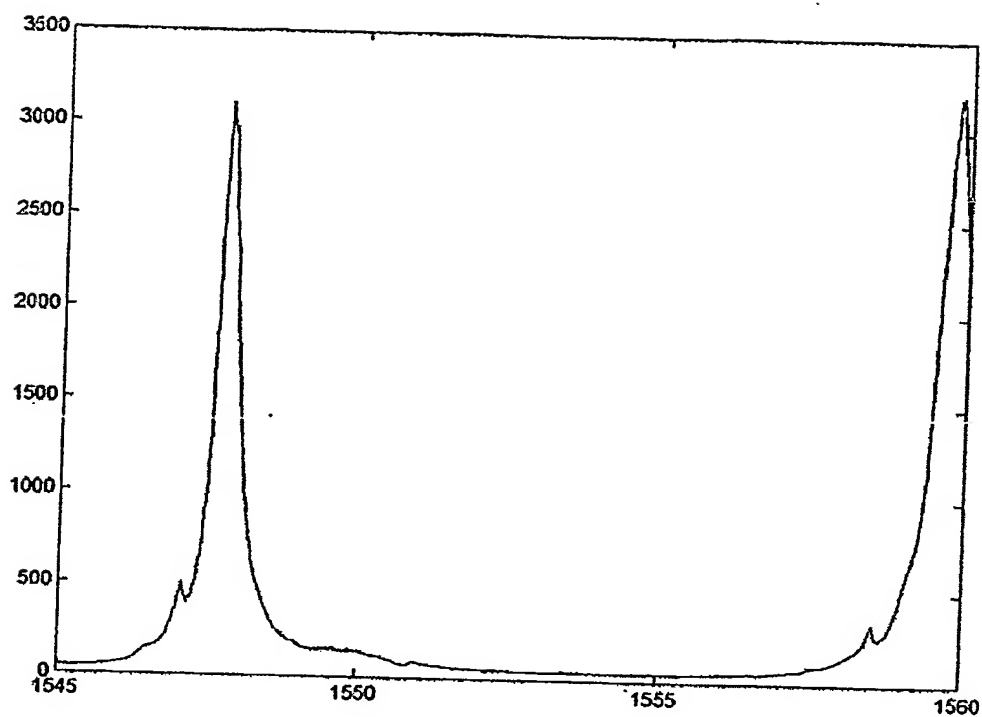


FIG 10D

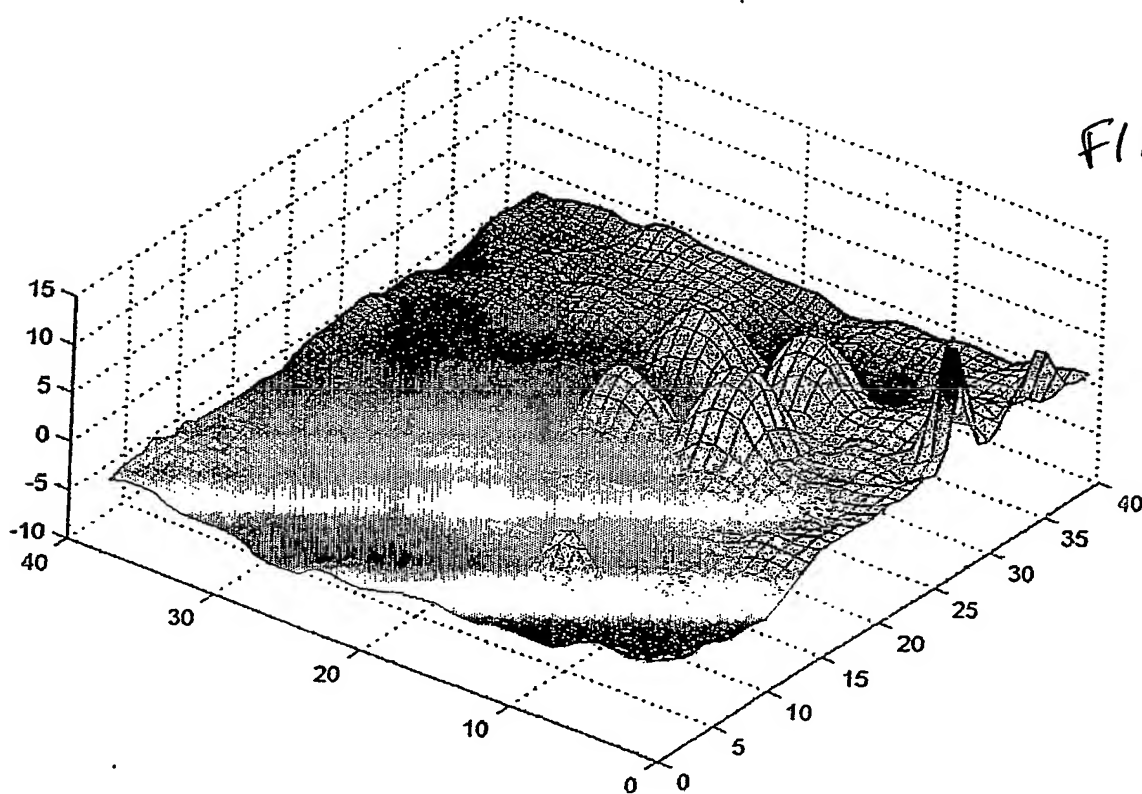


FIG 10E

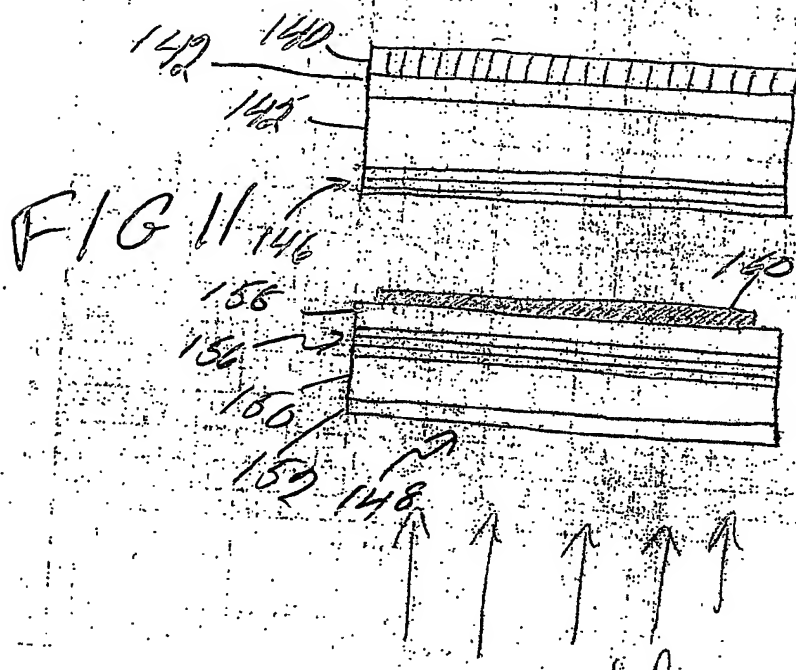


FIG 12.

